

Experience in extending the approach of cell-based TCP and MLD assays to Clostridium perfringens vaccines

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Novel *in-vitro* models as alternative to in-vivo toxoid vaccines testing: *Clostridium septicum* vaccine as proof of concept Virtual Workshop, 9 & 10 March 2021

Main objective within VAC2VAC project

 Contribute to the development of in vitro assays addressing specific toxicity of Clostridium perfringens, type C (noninactivated) antigen as an alternative to the currently used in vivo mouse tests.

Approach

- Use the same principle as described for in vitro toxicity testing of C. septicum vaccines. However, VERO cells are only poorly susceptible to the β-toxin of C. perfringens type C, considered the main antigen of this vaccine.
- Hence, main goals defined as follows:
 - **1.** Identification of cell line specifically susceptible to the *C. perfringens*, type C β -toxin.
 - **2.** In case of identification of such a cell line, show feasibility of assay development based on this cell line for assessment of C. perfringens C β -toxin activity in vitro.
 - ➤ Start with cell-based alternative to MLD in mice using *C. perfringens* C non-inactivated antigen
 - ➤ If successful, continue with development of cell-based assay (CBA) for assessment of residual toxicity of inactivated (toxoid) antigen

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First steps

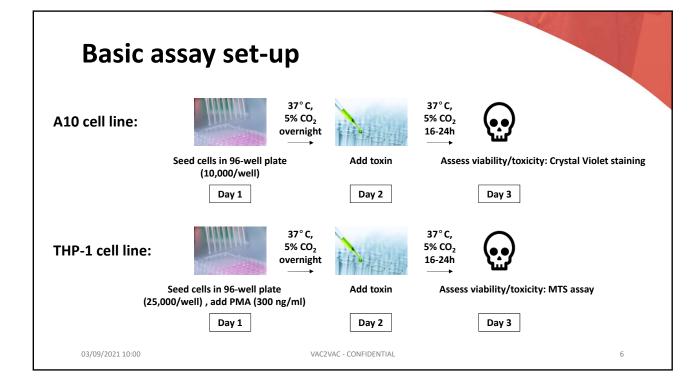
- Literature search for β-toxin-sensitive cell lines yielded the human HL-60 and THP-1 cell lines. In addition, one of our VAC2VAC industry partners recommended testing the rat A10 cell line.
- <u>Parallel approach:</u> Transfect VERO cells with P2X7 receptor, published to be the receptor involved in β -toxin activity (Nagahama *et al.*, 2015) to render the cells sensitive to β -toxin.

Initial results:

- HL-60 proved very difficult to culture and stable VERO-P2X7 transfectants did not show enhanced susceptibility to *C. perfringens* C non-inactivated antigen compared with parental VERO cells (Crystal Violet staining). → HL-60 & VERO-P2X7 not tested further!
- In contrast, **Rat A10 and human THP-1** showed β-toxin-induced toxicity in a concentration dependent manner using the MTS assay for THP-1 cells and Crystal Violet staining for the A10 cells.

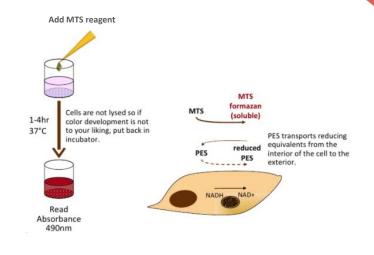
Next step

- Use of neutralizing Mab 10A2 against β -toxin and international anti- β -toxin standard to determine whether observed toxicity on **A10 and THP-1** cell lines is β -toxin-specific
 - > Crude *C. perfringens* C non-inactivated antigen used from one of our VAC2VAC industry partners in all experiments (also called 'end-of-fermentation supernatant'; contains β-toxin)
 - ➤ Mab 10A2 purchased from USDA
 - >International anti-β-toxin standard (CPBETAAT) purchased from NIBSC
 - ➤ Equal volumes of diluted non-inactivated antigen were pre-incubated with dilutions of antitoxin in medium, followed by 30 min incubation at 4°C on an orbital shaker (250 rpm).



MTS assay as a read-out for cell viability

- Assay is based on bioreduction of MTS by living cells into a soluble colored formazan product.
- The amount of formazan produced can be quantified by reading absorbance at 490nm.

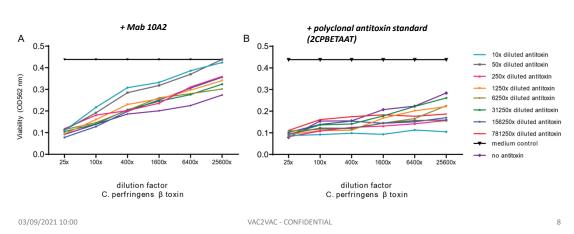


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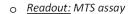
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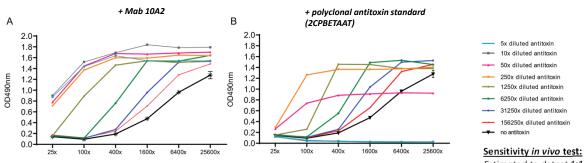
Toxicity of *C. perfringens* C non-inactivated antigen on A10 cells is only partially β-toxin dependent

o <u>Readout:</u> Crystal Violet staining



Toxicity of C. perfringens C non-inactivated antigen on THP-1 cells is β-toxin specific





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dilution factor

C. perfringens β-toxin

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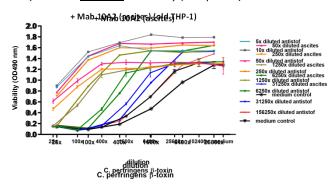
dilution factor

C. perfringens β toxin

Estimated to detect 1:100 -1:1000 dilution of same non-inactivated antigen material (VAC2VAC Industry partners, personal communication) 9

Results are reproducible with newly purchased THP-1 cells (ATCC: TIB-202)

PMA-treated THP-1 cells exposed to dilutions of C. perfringens non-inactivated antigen, either pre-incubated with anti-toxin (Mab 10A2) or not. Readout: MTS assay (in triplicate)



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Assay optimization - Cell density, PMA concentration & FBS amount

- Experimental set up used thus far:
 - 25.000 cells/well (96-well plates)
 - 300 ng/ml PMA
 - 10% FBS
- First optimization experiments showed that **50.000 cells/well** performed better than 25.000 or 100.000 cells/well (using 300 ng/ml PMA & 10% FBS; not shown).
- In addition, **75 ng/ml PMA** performed better than 150 or 300 ng/ml (using 25.000 cells/well & 10% FBS; not shown).

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Assay optimization - Cell density & PMA concentration

Dilution CPB-toxin

→ 75 ng/mL PMA → 50 ng/mL PMA → 25 ng/mL PMA

Dilution CPB-toxin

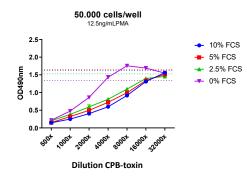
Best performing conditions: 50.000 cells/well & 12.5 ng/ml PMA

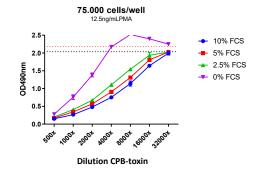
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Dilution CPB-toxin

Assay optimization - Cell density & FBS amount



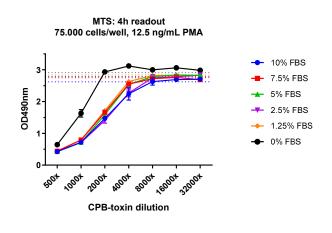


Best performing conditions:

75.000 cells/well & 12.5 ng/ml PMA
2.5, 5 or 10% FBS does not seem to make a large difference

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Assay optimization - FBS amount



Conclusions:

1.25% - 10% FBS has only minor influence on the assay.

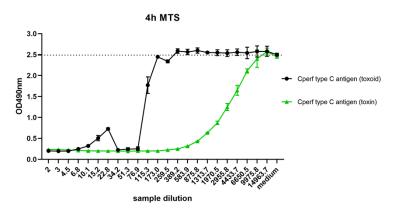
0% FBS: Curve reaches plateau already at relatively low dilutions. Suggests that assay has less discriminatory power when no FBS is used.

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Further development THP-1 CBA to assess toxicity of *C. perfringens* C inactivated antigen (toxoid)



- · Dotted line represents the average absorbance reading of untreated control wells
- 1.5-fold dilution series

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Conclusions (I)

- A10 cells are susceptible to *C. perfringens* type C non-inactivated antigen but observed toxicity is not β-toxin specific
- PMA-treated THP-1 cells are specifically susceptible to the β-toxin that is present in the *C. perfringens* type C non-inactivated antigen
- Results with the 'in-house' THP-1 cells can be reproduced with 'new' THP-1 cells ordered at ATCC
- Optimized experimental setup includes 75,000 cells per well (96-well plate) and 12.5 ng/ml PMA, in which the amount of FBS can be anywhere between 1.25% and 10%
 - ➤ An optimized protocol for the THP-1-based MLD assay has been prepared and transferred to the industry partners for further assessment and validation

Conclusions (II)

- THP-1 CBA may be suitable for assessment of residual toxicity of inactivated (toxoided) antigen preparations. However, there is a problem to solve:
 - The inactivated antigen still contains residual formaldehyde. This makes it difficult to determine whether the cell death observed at low dilutions is caused by residual β-toxin or by formaldehyde (or both).
 - **Next step:** Repeat the THP-1 CBA with inactivated antigen in the presence of formaldehyde-neutralizing agents

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